CLIMATE CHANGE: MOLECULAR ADAPTATION STRATEGIES IN CEREALS AND FORAGE CROPS-A REVIEW

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SUMMARY

Owing to various anthropogenic activities, atmospheric CO_2 concentration continues to rise and in turn enhance average global temperature causing climate warming. Elevated atmospheric CO_2 , enhances Ribulose1,5-bisphosphate carboxylase (photosynthesis) activity and reduces Ribulose1,5-bisphosphate oxygenase activity (photorespiration) in C_3 plants. Annual mean temperature has increased about 0.4 °C over the last century. Increased temperature affects crop yield, as it shortens plant growth and developmental phases. With continued increase in population, crop yields must also increase to meet the future requirements of food and fodder. This article reviews the impact of elevated CO_2 and temperature on physiological and biochemical responses of cereals and forage crops and also the molecular adaptation strategies for the development of climateresilient crops for sustainable food and fodder security.

Key words: Climate change, molecular adaptation mechanism, cereals, forage crops

The climate emergency owing to the increased emission of greenhouse gases and other anthropogenic activities is posing risk to sustainable food and fodder security worldwide. Atmospheric CO₂ concentration has elevated from approximately 280 ppm in the preindustrial era to 400 ppm presently, due to excessive use of fossil fuels and other activities including deforestation. The average global temperature increased by about 1°C since 1880 (Crous, 2019). Atmospheric CO, concentrations continue to rise and is predicted to reach approximately 550 ppm by 2050 and 700 ppm by the end of this century (Becklin et al., 2016). This would further enhance warming with average global temperature predicted to be increased by 1.1-4.8 °C depending on the magnitude of the emissions of greenhouse gases (IPCC, 2014; Borland 2015). Further, high latitudes may experience severe warming, about 10°C increase in temperature, by the year 2100, on the other hand, the tropics may experience comparatively smaller temperature increases of about 3-4°C (Ciais *et al.*, 2013). Furthermore, climate warming is expected to cause changes in the precipitation patterns leading to extremes in dry and wet spells (Medvigy and Beaulieu, 2012).

Climate warming is predicted to have enormous impact on the Earth's flora and fauna. The continued rise in global CO₂ levels and the climate change phenomenon thereupon will have huge influence on ecosystems and other natural resources. Agriculture and allied activities are predicted to be most affected as climate warming will have direct as well as indirect effect on crop production, soil microbes, livestock and pests. The predicted extreme changes in the climate may detrimentally affect crop productivity. Furthermore, human population is ever growing and have been predicted to reach 9 billion by

2050 necessitating approximately 70 % enhancement in crop production (FAOSTAT, 2017; Bhat *et al.*, 2020). Therefore, ensuring food, fodder and nutritional security and development of crop varieties with high nutritional and yield potential and are adaptable to the challenging future climate scenario is of prime importance.

Cereals including wheat, rice, maize, barley and millets serve as staple crops and are among the major food grains produced across the globe. The pros and cons of climate change on cereal crops will have a tremendous influence on food security as they are among the staple foods. There is a high demand for cereal and forage yield and seed production at the present and in future as well. A major challenge ahead for those involved in the seed industry, therefore, is to provide cultivars that can maximize future crop yields under the changing climate (Ainsworth *et al.*, 2008b; Bruins 2009; Ceccarelli et al., 2010). Climate resilient crops such as leguminous forages: berseem, cowpea, centro, clitoria and siratro (Dheeravathu et al., 2017a and b, Dheeravathu et al., 2021a, Dheeravathu et al., 2021c, Dheeravathu et al., 2022), grasses: guinea grass, bajra-Napier hybrids and a tri-specific hybrid, dinanath grass (Dheeravathu et al., 2018, Singh et al., 2020, Dheeravathu et al., 2021b, Singh et al., 2021, Antony et al., 2021, Dheeravathu et al., 2022), cereal forage: oat (Dheeravathu et al., 2022), and forage millets: sorghum, pearl millet (Singh et al., 2010) have been proven to be climate smart. Considering the effect of elevated CO₂ and temperature on forage growth, yield and productivity, identification of physiological and biochemical changes mechanisms and molecular adaptation strategies in cereal and forage crop varieties/genotypes/lines could play a major role in sustaining food, forage and livestock production and will be helpful in future breeding programs. In this review, the impact of elevated CO₂ and temperature on cereal and forage crops and also the various strategies adaptation to tackle the detrimental effects for sustainable agriculture and food security are discussed.

Effect of rising CO₂ and elevated temperature on cereals and forage crops

Rising CO₂ levels directly impact plant metabolism owing to its role in the photosynthetic process and hence are expected to evoke profound physiological and biochemical alterations in plants. Increased CO₂ indirectly impacts plant performance

through its effect on temperature and water stress as well as by directly affecting the plant metabolism through its role in photosynthesis (Kanwal et al., 2014; Abebe et al., 2016). Under elevated atmospheric CO₂, Ribulose1,5-bisphosphate carboxylase/oxygenase (Rubisco), a key enzyme of photosynthetic pathway, can better bind to CO₂, enabling enhanced carbon fixation. Thus, in general, elevated CO, levels stimulate photosynthesis initially and also simultaneously supress oxygenation that causes photorespiration (Bowes 1991). The enhancement of photosynthetic process due to elevated CO₂ is profound only when the internal CO, concentration is low; i.e., when the rate of photosynthesis is limited by rubisco carboxylation. With further increase in the internal CO, concentration, the photosynthetic process becomes less dependent of CO₂ concentration (as it already saturated) and is limited by the ability to regenerate RuBP. Thus, increase in the CO, levels in C, plants (including rice and wheat among others) results in net stimulation of photosynthesis initially and hence increased biomass accumulation (Ainsworth and Long, 2005). Improved tillering and shoot biomass have been reported in wheat under increased CO₂ levels (Bourgault et al., 2013). But with further increase in CO, concentration for longer periods, stimulation of photosynthesis is not maintained due to acclimation process (Long et al., 2004). On the other hand, plants with C₄ photosynthetic pathway (including maize, sorghum, pearl millet) devise a different primary carboxylase, phosphoenolpyruvate carboxylase (PEP carboxylase). PEP carboxylase assimilates CO,, fixes it as oxaloacetate in the mesophyll cells which is then decarboxylated in the bundle sheath cells. Thus, in C₄ plants, CO, is delivered to the bundle sheath cell rubisco in higher concentrations enhancing photosynthetic efficiency and inhibiting photorespiration (Leakey, 2009; Kant et al., 2012). Rise in atmospheric CO, levels will lead to a rise in intracellular CO₂ concentration in C₄ plants; however, it may not have profound effect on carbon fixation and biomass production as they already are nearly photosynthetically saturated (Ainsworth and Long, 2005; Kant et al., 2012). Free-air CO, enrichment studies revealed enhanced yield in C3 plants under elevated CO, without any stress on the other hand, productivity C₄ plants were not increased except under drought stress (reviewed in Ainsworth and long, 2020). Nevertheless, significant enhancement in yield and biomass production of C₄ nutri cereal, foxtail millet was observed under elevated CO, levels as determined

by open-top chamber studies (Gong *et al.*, 2021). For other growth and/yield related characteristics, plant responses to elevated CO₂ levels vary considerably (Kadam *et al.*, 2015).

Further, rise in CO, levels considerably lowers stomatal conductance and in general improves the water use efficiency (WUE), as have been reported in wheat (Li et al., 2004). Decreases up to 20% stomatal conductance have been reported in the Free Air CO, Enrichment (FACE) owing to rise in CO₂ levels, while, in some studies (Pathare et al., 2017), lowering of stomatal conductance have not been observed. Nevertheless, the decreased stomatal conductance leads to increased resistance in CO₂ diffusion and thus offsets rubisco carboxylation rates. Lowered stomatal conductance enhances soil water savings, enabling to compensate the expected future higher evaporative demand thus promoting crop productivity. Furthermore, reduced stomatal conductance and the lowered transpiration rates results in elevated leaf temperature (Madan et al., 2012) which in turn causes greater leaf to air vapour pressure difference (VPD), a driving force for transpiration. This counterbalances the water savings from decreased stomatal conductance under elevated CO₂ and hence the overall transpiration rate under elevated CO₂ is only minimum as compared to ambient CO₂.

Effect of elevated CO₂ on flowering time shows that there exist slight advancements in flowering time in many C, plants including cereals viz. rice and barley (Craufurd and wheeler, 2009). Also, higher spikelet sterility was observed in rice and sorghum owing to enhanced CO, during anthesis (Matsui et al., 1997). Further, elevated CO, induced decrease in grain protein and micronutrients have been reported in some crops; for instance, Li et al. (2019) found a decrease in grain Ca and K content in wheat. More recently, Gong et al. (2021) reported that elevated CO₂ decreased Fe content of foxtail millet grains while it enhanced accumulation of P in the grains. As temperature plays critical role in all the processes associated with plant growth and development, elevated temperature would have profound impact on crop productivity. Increased temperature affects crop yield, as it shortens plant growth and developmental phases through alterations in the rate and timing of physiological processes (Ahad and Reshi, 2015, Dheeravathu et al., 2022). Enhanced leaf temperatures result in increased stimulation of photorespiration owing to the shift in specificity of Rubsico for O₂ at higher temperatures and also enhanced solubility of O, as compared to CO₂ in turn leading to higher availability of O₂.

Elevated temperature beyond the optimum temperature for photosynthesis would have negative effect on plant growth and yield. The temperature optimum of photosynthesis may vary among plant species with those adapted to hotter climates having higher values as compared to those adapted to colder region (Sage and Kubien, 2007). Also, C₄ plants possess a higher optimum temperature as compared to C, plants (Crafts-Brander and Salvucci, 2002). Decline in photosynthesis at elevated temperatures (i.e., temperatures above the thermal optimum of assimilation) can be explained in terms of heat labile nature of enzymes including rubisco activase, the enzyme that catalyses removal of inhibitory molecules from the catalytic site of rubisco and the reduction in the electron transport at elevated temperatures (Dusenge et al., 2019). Lower shoot biomass accumulation during vegetative phase owing to decreased photoassimilate production caused by heat stress induced on photosynthetic machinery have been reported in maize (Sinasawat et al., 2004). Further, rise in air temperature causes an increase in the air saturation vapour pressure leading to an increased vapour pressure deficit between air and leaf which in turn decrease water-use efficiency of plants (Ahad and Reshi, 2015). High temperature stress adversely affects reproductive stages of crop development in cereals often causing fewer spikelets/grains and reduced sink size (Kadam et al., 2014). Elevated temperatures above the optimal range adversely affects pollination process; heat stress induced floral abnormalities including pistil hyperplasia adversely affects reproduction in rice (Takekova et al., 1991). Reduced pollen viability, poor pollen germination, anther indehiscence induced by heat stress have been reported in rice (Jagadish et al., 2007; Rang et al., 2011). High temperature stress during pre-anthesis stage decreases pollen or ovule viability and/or stigma receptivity (Prasad et al., 2008, Nguyen et al., 2013, Djanaguiraman et al., 2014, Prasad and Djanaguiraman, 2014), causes reproductive structure abnormalities (Prasad and Djanaguiraman, 2014) and oxidative damage, resulting in pollen sterility and decreased seed-set (Djanaguiraman et al., 2014). High temperature stress at the time of anthesis can decrease floret fertility even when the pollen is viable (Prasad and Djanaguiraman, 2014). The other negative impacts of elevated temperatures in cereal crops include drastic reduction in grain yield. Low grain weight in cereals including rice and wheat associated with poor assimilate remobilization and poor grain filling due to increased temperature have been observed (Kadam et

al., 2014). Furthermore, heat stress impacts antioxidant enzymes; for instance, negative influence of heat stress on antioxidant enzymes of maize have been reported by Gong *et al.* (1997).

Climate change: Application of genomics and molecular breeding approaches in the molecular adaptation mechanism in cereals and forage crops

The phenomenon of global climate change, with its wide spread impact on agricultural productivity, continues to be one of the major environmental challenges of twenty-first century. With the future predictions of extreme weather events, owing to high carbon emissions, it is implicit that better strategies to adapt to the changing climate scenario are needed. Cultural methods *viz*. alterations in the crop planting and harvesting time, changes in the cropping schemes, crop rotation and adoption of crops with shorter life cycle *etc* are devised by farmers. With the prediction of extreme heat in the coming years, breeding efforts to develop cultivars that are adapted to heat stress, drought and other abiotic stress would enable sustainable crop improvement and productivity.

Conventional breeding approaches are valuable tools in developing stress-resistant crop cultivars; however, more efficient and rapid strategies are needed to address present climate change and food insecurity challenges. Crop improvement strategies are based upon genetic diversity of crop species and the development of crop cultivars with improved adaptation to various abiotic stress would be beneficial in mitigating the effects of climate change. Land races of crops serve as important source of genetic diversity as they display adaptation to agro-climatic conditions in which they survive and are hence excellent tools for crop improvement programs (Lopes et al., 2015). Evaluation of drought response in 105 land races of pearl millet on a wide range of environmental conditions enabled the identification of 15 land races with high degree of tolerance to drought that can be used in the development of drought tolerant cultivars (Yadav et al., 2003). With the advancement in molecular biology tools, genotype-based strategies started gaining importance. Advanced tools employed for the development of improved crop cultivars include genetic engineering (GE)-based strategies and molecular breeding approaches (Kole et al., 2015). GE based strategies allow transfer of gene (s) of interest to crop plants for generating desired phenotype. This technique of transgenic plant development has been widely employed to develop crops that are resistant to a number of abiotic stresses. For instance, transgenic rice, over expressing *OsDREB2A* under a stress inducible promoter from *Arabidopsis* with improved resistance to salinity and drought have been developed (Mallikarjuna *et al.*, 2011). Further, transgenics enabled better understanding of various aspects of plant development including response to stresses. Nevertheless, the integration of foreign genes into edible crops are controversial in many countries (Kole *et al.*, 2015; Jaganathan *et al.*, 2018).

Molecular breeding approaches are promising as they enable expansion of the size of breeding program, in turn enhancing selection intensity (Yuan et al., 2019). DNA markers or molecular markers including RFLP, AFLP, SSRs, SNPs etc have been widely employed to identify QTLs for understanding the underlying mechanism of stress adaptation in crops including cereals and thereby contributing to crop improvement under stress condition. The markerassisted selection (MAS) for various crop improvement strategies including those targeting abiotic stress tolerance have accelerated breeding strategies with reduced need for extensive field selection. Linkage mapping to identify markers associated with traits of interest have been widely employed in several crops including cereals (Kulwal et al., 2011). Yadav et al. (2002) performed QTL mapping to dissect traits determining grain and stover yield under terminal drought tolerance in pearl millet. However, attempts to improve complex traits by QTL -associated markers displayed limited success owing to the low resolution in the identification of markers associated with small effect QTLs (Kulwal et al., 2011) and also difficulty of finding same QTLs across different environments (Crossa et al., 2017). Hence, association mapping with its increased mapping resolution and greater allele number is being widely employed for dissecting complex traits in crops.

The recent advances in genomics and allied areas, allowed molecular breeding approaches to further comprehensively characterize allelic diversity underlying agronomic traits of interest including those confer abiotic stress tolerance and their application in the development of novel cultivars with improved climate resilience. Advances in NGS offers cost effective genotyping assays, large numbers of numerous genomic resources including genome-wide molecular markers *viz*. SNPs and insertion-deletions (InDels) *etc* have been developed for crops which are serving as promising tools for genomics assisted

breeding (Kole et al., 2015). Owing to their high density, SNPs are among the markers of choice for QTL mapping. Recently, the rapid and robust technique of Genotyping by sequencing (GBS) have been widely employed for the generation of genome wide SNPs (Poland and Rife, 2012). High-density linkage map construction and QTL analysis in maize, wheat, barley and oats were made possible through GBS (Poland et al., 2012; Huang et al., 2014). GBS also enabled the identification of alleles or genes responsible for the trait of interest in cereal crops including rice for seedling salinity stress tolerance (De Leon et al., 2016), pearl millet for drought tolerance (Debieu et al., 2018). These molecular markers are widely being used to assess genetic diversity, population structure and also to dissect genomic regions associated traits of interest through genome-wide association study (GWAS). GWAS is a powerful tool widely employed for the detection of genomic regions associated with any given trait and determination of statistical association between genetic polymorphisms including SNPs and trait variations in large germplasm collections. GWAS have been employed for the identification of candidate genes associated with agronomic traits including grain yield and the secondary traits under well-watered and drought stress conditions in maize (Xue et al., 2013; Thirunavukkarasu et al., 2014). Also, through GWAS, 365 single nucleotide polymorphisms (SNPs) associated with drought-related traits in maize was identified (Li et al., 2016).

Furthermore, the high throughput genotyping and phenotyping techniques enables the employment of genomic selection approach. Genomic selection is a promising tool that enables to improve complex traits by capturing both major small effect QTLs with high density genome-wide marker coverage. Genomic selection increases the genetic gain per selection thus enhancing efficiency of breeding programs and is considered as an excellent alternative to MAS and other strategies (Srivastava et al., 2020). Genomic selection studies have been employed in the cereal crops including wheat, maize (Bevene et al., 2015; Shikha et al., 2017) and pearl millet (Varshney et al., 2017). Also, more recently, genome editing has emerged as an important tool to manipulate crop genome for better crop performance under biotic and abiotic stress conditions (Kole et al., 2015). This strategy utilizes sequence specific nucleases and is a promising alternative to conventional breeding and incorporation of foreign DNA under the current climate change scenario. Zinc finger nucleases (ZFNs), Engineered homing endonucleases/meganucleases (EMNs), transcription activator-like effector nucleases (TALENs) and clustered regularly interspaced short palindromic repeats/CRISPR-associated 9 (CRISPR/Cas9) are among the widely employed genome editing strategies. (Townsend *et al.*, 2009; Silva *et al.*, 2011; Cermak *et al.*, 2011). The application of CRISPR/Cas tool for crop improvement under abiotic stresses have been employed in cereal plants including rice (Zhang *et al.*, 2014; Shan *et al.*, 2014) and maize (Shi *et al.*, 2017).

CONCLUSION

The continued rise in global CO, levels and the climate change phenomenon thereupon will have huge influence on forage and cereal crop production and productivity. Increased temperature and drought stress incidences due to climate change reduce the quality and quantity of pollen grains resulting in low seed formation causing significant loss of forage and crop yield. Development of forage and crop adaptive traits with high nutritional and superior yield potential are recommended to meet the challenging future climate scenario to ensure food and nutritional security for the burgeoning global population. The advent of high throughput genotyping and phenotyping platforms with advanced genomic tools including the genome wide association studies, gene editing and genomic selection would enable better understanding of molecular mechanism of response to various stress and breeding of climate resilient cereal and forage crops. Knowledge on molecular mechanisms of plant adaptation to extreme temperatures/ heat stress, drought and other abiotic stresses are essential for effective management of unfavourable agro-ecologies in the wake of climate change. Despite great efforts in understanding the molecular mechanism associated with stress tolerance, more research is needed on molecular adaptation strategies on cereals and forage crops.

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REFERENCES

- Ahad, B. and Z. Reshi, 2015: Climate Change and Plants. In: Crop Production and Global Environmental Issues (eds. Hakeem, K.) Springer, Cham. https://doi.org/10.1007/978-3-319-23162-4 20.
- Abebe, A., H. Pathak, S. D. Singh, A. Bhatia, R. C. Harit and V. Kumar. 2016: Growth, Yield, and Quality of Maize with Elevated Atmospheric Carbon Dioxide and Temperature in North-west India. *Agric. Ecosyst. Environ.*, 218: 66-72.
- Ainsworth, E. A. and S. P. Long, 2020: 30 years of free-air carbon dioxide enrichment (face): what have we learned about future crop productivity and its potential for adaptation? *Global Change Biol.*, 27:1-23.
- Ainsworth, E. A., C. Beier, C. Calfapietra, R. Ceulemans, M. Durand-Tardif, G. D. Farquhar, D. L. Godbold, G. R. Hendrey, T. Hickler, J. Kaduk, D. F. Karnosky, B. A. Kimball, C. Körner, M. Koornneef, T. Lafarge, A. D. Leakey, K. F. Lewin, S. P. Long, R. Manderscheid, D. L. Mcneil, T. A. Mies, F. Miglietta, J. A. Morgan, J. Nagy, R. J. Norby, R. M. Norton, K. E. Percy, A. Rogers, J. F. Soussana, M. Stitt, H. J. Weigel, and J. W. White, 2008b: Next generation of elevated CO₂ experiments with crops: a critical investment for feeding the future world. *Plant Cell Environ*. 31: 1317-1324.
- Ainsworth, E. A. and S. P. Long, 2005: What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. *New Phytol.* **165**: 351-372.
- Becklin, K. M., J. T. Anderson, L. M. Gerhart, S. M. Wadgymar, C. A. Wessinger, and J. K. Ward, 2016: Examining Plant Physiological Responses to Climate Change through an Evolutionary Lens. *Plant Physiol.* 172: 635-649.
- Beyene, Y., K. Semagn, S. Mugo, A. Tarekegne, R. Babu, and B. Meisel, et al. 2015: Genetic gains in grain yield through genomic selection in eight biparental maize populations under drought stress. *Crop Sci.* 55: 154-163.
- Bhat, J. A., R. Deshmukh, T. Zhao, G. Patil, A. Deokar, S. Shinde, and J. Chaudhary, 2020: Harnessing High-throughput Phenotyping and Genotyping for Enhanced Drought Tolerance in Crop Plants. *J. Biotechnol.* **324**: 248-260.
- Borland, A. M., S. D. Wullschleger, D. J. Weston, J. Hartwell, G. A. Tuskan, X. Yang, and J. C. Cushman, 2015: Climate-resilient agroforestry: physiological responses to climate change and engineering of crassulacean acid metabolism (CAM) as a mitigation strategy. *Plant Cell Environ.* **38**:1833-1849.

- Bourgault, M., M. F. Dreccer, A. T. James, and S. C. Chapman, 2013: Genotypic variability in the response to elevated CO2 of wheat lines differing in adaptive traits. *Funct. Plant Biol.* **40**: 172-184.
- Bowes, G. 1991: Growth at elevated CO₂: photosynthetic responses mediated through Rubisco. *Plant Cell Environ.* **14**: 795-806.
- Bruins, M., 2009: The evolution and contribution of plant breeding to global agriculture. In Proceedings of the Second World Seed Conference: Responding to the Challenges of a Changing World: The Role of New Plant Varieties and High-Quality Seed in Agriculture, 18-31.
- Ceccarelli, S., S. Grando, M. Maatougui, M. Michael, M. Slash, R. Haghparast, M. Rahmanian, A. Taheri, A. Alyassin, A. Benbelkacem, M. Labdi, H. Mimoun, and M. Nachit, 2010: Plant breeding and climate changes. *Journal of Agricultural Science, Cambridge* 148: 627-637.
- Cermak, T., E. L. Doyle, M. Christian, L. Wang, Y. Zhang, and C. Schmidt, et al. 2011: Efficient design and assembly of custom TALEN and other TAL effector-based constructs for DNA targeting. *Nucleic Acids Res.* **39**: e 82.
- Ciais, P., C. Sabine, G. Bala, L. Bopp, V. Brovkin, J. Canadell, A. Chhabra, R. DeFries, et al. 2013: Carbon and other biogeochemical cycles. In: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (eds Tignor, M., Allen, S. K., Boschung, J., Nauels, A., Xia, Y., Bex, V., Midgley, P.M.) Cambridge University Press, Cambridge, UK and New York, NY, USA, 465-570.
- Crafts-Brandner, S. J. and M. E. Salvucci, 2002: Sensitivity of photosynthesis in a C₄ plant, maize, to heat stress. *Plant Physiol.* **129**: 1773-1780.
- Craufurd, P. Q. and T. R. Wheeler, 2009: Climate change and the flowering time of annual crops. *J. Exp. Bot.* **60**: 2529-2539.
- Crossa, J., P. Pérez-Rodríguez, J. Cuevas, O. Montesinos-López, D. Jarquín, et al. 2017 : Genomic Selection in Plant Breeding: Methods, Models, and Perspectives. *Trends Plant Sci.* 22 : 961-975.
- Crous, K. Y. 2019: Plant responses to climate warming: physiological adjustments and implications for plant functioning in a future, warmer world. *Am. J. Bot.* **106**: 1049-1051.
- De Leon, T. B., S. Linscombe, and P. K. Subudhi, 2016: Molecular Dissection of Seedling Salinity Tolerance in Rice (*Oryza sativa L.*) Using a High-Density GBS-Based SNP Linkage Map. *Rice* 9: 52.
- Debieu, M., B. Sine, S. Passot, A. Grondin, E. Akata, and P. Gangashetty, et al. 2018: Response to early

- drought stress and identification of QTLs controlling biomass production under drought in pearl millet. *PLoS One* **13**: e 0201635.
- Dheeravathu, S. N., T. B. Vadithe, N. Dikshit, T. N. Usha, S. N. Vadithe, R. Venkateswarlu, V. Manasa, and G. S. Bandeppa, 2022: Effect of elevated CO2 and temperature on physiological and biochemical changes in forage crops A review. Forage Res. 48: 22-27.
- Dheeravathu, S. N., P. Singh, R. Srinivasan, and V. K. Yadav, 2022: Open Top Chamber: An innovative screening technique for temperature stress tolerance in forage oat (*Avena sativa*). Forage Res. 47: 513-516.
- Dheeravathu, S. N., T. Singh, A. Radhakrishna, Reetu, R. Gajghate, S. R. Kantwa, and H. A. Bhargavi, 2021a: Effect of salinity stress on different seed vigour indices in single and multicut berseem (*Trifolium alexandrinum*) varieties. *Forage Res.* 46: 368-373.
- Dheeravathu, S. N., K. Singh, P. K. Ramteke, Reetu, N. Dikshit, M. Prasad, D. Deb, and T. B. Vadithe, 2021b: Physiological responses of Bajra-Napier hybrids and a tri-specific hybrid to salinity stress. *Trop. Grassl. Forrajes Trop.* 9: 337-347.
- Dheeravathu S. N., M. H. Hanamant, T. B. Vadithe, S. N. Vadithe, K. Singh, N. Dikshit, T. N. Usha, T. Singh, Reetu and R. Gajghate, 2021c: Salinity tolerance of forage cowpea [Vigna unguiculata (L.) walp.] during germination and early seedling growth. Forage Res. 47: 213-221.
- Dheeravathu, S. N., V. C. Tyagi, C. K. Gupta and A. Edna, 2018: Manual on Plant Stress Physiology. ICAR Indian Grassland and Fodder Research Institute, Jhansi. Stress assessment formulas and stress related terminology: 1-87.
- Dheeravathu, S. N., A. Edna, R. V. Koti, and M. B. Doddamani, 2017a: Salinity tolerance of forage range legumes during germination and early seedling growth. *Progressive Res. J.* 12: 1357-1360.
- Dheeravathu S. N., T. Singh and A. Radhakrishna, 2017b: Effect of drought stress on biomass and drought adaptive traits in Berseem (*Trifolium alexandrinum* L.), National symposium-new directions in managing forage resources and livestock productivity in 21st century: challenges and opportunity: 4-17.
- Djanaguiraman, M., P. V. V. Prasad, M. Murugan, R. Perumal, and U. K. Umesh, 2014: Physiological differences among sorghum (Sorghum bicolor L.Moench) genotypes under high temperature stress. Environ. Exp. Bot. 100: 43-54.
- Dusenge, M. E., A. G. Duarte, and D. A. Way, 2019: Plant carbon metabolism and climate change: elevated CO, and temperature impacts on photosynthesis,

- photorespiration and respiration. *New Phytol.* **221**: 32-49.
- Edna, A., A. B. Kawadikai, S. Hullur, K. Sridhar, S. Nayak, and V. K. Yadav, 2021: Biomass repartitioning, tiller regeneration and salt secretion through leaf micro hairs for salinity tolerance in guinea grass (Megathyrsus maximus Jacq.) *Range Mgmt and Agroforestry* **42**: 246-254.
- FAOSTAT, 2017: Available online: http://www.fao.org/faostat/en/#data.
- Gong, M., S. N. Chen, Y. Q. Song, and Z. G. Li, 1997: Effect of calcium and calmodulin on intrinsic heat tolerance in relation to antioxidant systems in maize seedlings. *Funct. Plant Biol.* 24: 371-379.
- Gong, Z., L. Dong, S. Lam, D. Zhang, Y. Zong, X. Hao, and Li, P. 2021: Nutritional quality in response to elevated CO₂ concentration in foxtail millet (Setaria italica). J. Cereal Sci. 102: 103318.
- Huang, Y. F., J. A. Poland, C. P. Wight, E. W. Jackson, and N. A. Tinker, 2014: Using genotyping-by-sequencing (GBS) for genomic discovery in cultivated oat. *PLoS ONE* 9: e102448.
- IPCC, 2014: Climate change 2014: impacts, adaptation, and vulnerability. In Contribution of the Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (eds Field, C., Barros, V., Mach, K., Mastrandrea, M.), Cambridge University Press, Cambridge, UK/New York, NY, USA.
- Jagadish, S. K. V., P. Q. Craufurd, and T. R. Wheeler, 2007 : High temperature stress and spikelet fertility in rice (*Oryza sativa* L.). *J. Exp. Bot.* **58**: 1627-1635.
- Jaganathan, D., K. Ramasamy, G. Sellamuthu, S. Jayabalan, and G. Venkataraman, 2018: CRISPR for Crop Improvement: An Update Review. Front. Plant Sci. 9: 985.
- Kadam, N. N., G. Xiao, R. J. Melgar, R. N. Bahuguna, and C. Quinones, et al. 2014: Chapter Three Agronomic and Physiological Responses to High Temperature, Drought, and Elevated CO₂ Interactions in Cereals, In *Advances in Agronomy* (eds Donald Sparks) Academic Press 127: 111-156.
- Kant, S., S. Seneweera, J. Rodin, M. Materne, D. Burch, S. J. Rothstein, and G. Spangenberg, 2012: Improving yield potential in crops under elevated CO₂: integrating the photosynthetic and nitrogen utilization efficiencies. *Front. Plant Sci.* **3**:162.
- Kole, C., M. Muthamilarasan, R. Henry, D. Edwards, and R. Sharma, et al. 2015: Application of genomics-assisted breeding for generation of climate resilient crops: progress and prospects. *Front. Plant Sci.* **6**: 563.
- Kanwal, P., S. Gupta, S. Arora, and A. Kumar, 2014: Identification of genes involved in carbon

- metabolism from *Eleusine coracana* (L.) for understanding their light-mediated entrainment and regulation. *Plant Cell Rep.* **33**: 1403-1411.
- Kulwal, P., T. Mahendar, and R. K. Varshney, 2011: Genomics interventions in crop breeding for sustainable agriculture. In: Encyclopedia of Sustainability Science and Technology (eds Robert E Meyers) Springer.
- Leakey, A. D. 2009: Rising atmospheric carbon dioxide concentration and the future of C4 crops for food and fuel. *Proc. Biol. Sci.* **276**: 2333-2343.
- Li, C., B. Sun, and Y. Li, et al. 2016: Numerous genetic loci identified for drought tolerance in the maize nested association mapping populations. *BMC Genomics* 17: 894.
- Li, F., S. Kang, and J. Zhang, 2004: Interactive effects of elevated CO₂, nitrogen and drought on leaf area, stomatal conductance, and evapotranspiration of wheat. *Agr. Water Manage.* **67**: 221-233.
- Li, X., A. Khan, Z. Lv, L. Fang, D. Jiang, F. Liu, 2019: Effect of multigenerational exposure to elevated atmospheric CO₂ concentration on grain quality in wheat. *Environ. Exp. Bot.* **157**: 310-319.
- Long, S. P., E. A. Ainsworth, A. Rogers, and D. R. Ort, 2004: Rising atmospheric carbon dioxide: plants FACE the future. *Annu Rev Plant Biol.* 55: 591-628
- Lopes, M. S., I. El-Basyoni, P. S. Baenziger, S. Singh, C. Royo, K. Ozbek, H. Aktas, E. Ozer, F. Ozdemir, A. Manickavelu, T. Ban, and P. Vikram, 2015: Exploiting genetic diversity from landraces in wheat breeding for adaptation to climate change. *J. Exp. Bot.* 66: 3477-3486.
- Madan, P., S. V. K. Jagadish, P. Q. Craufurd, M. Fitzgerald, T. Lafarge, and T. R. Wheeler, 2012: Effect of elevated CO₂ and high temperature on seed-set and grain quality of rice. *J. Exp. Bot.* **63**: 3843-3852.
- Mallikarjuna, G., K. Mallikarjuna, and M. K. Reddy, 2011
 : Expression of OsDREB2A transcription factor confers enhanced dehydration and salt stress tolerance in rice (*Oryza sativa* L.). *Biotechnol. Lett.* **33**: 1689-1697.
- Matsui, T., O. S. Namuco, L. H. Ziska, and T. Horie, 1997:
 Effects of high temperature and CO₂
 concentration on spikelet sterility in indica rice.
 Field Crops Res. 51: 213-219.
- Medvigy, D. and C. Beaulieu, 2012: Trends in daily solar radiation and precipitation coefficients of variation since 1984. *J. Clim.* **25**: 1330-1339.
- Nguyen, C. T., V. Singh, E. J. van Oosterom, S. C. Chapman, D. R. Jordan, and G. L. Hammer 2013: Genetic variability in high temperature effects on seed set in sorghum. *Funct. Plant Biol.* 40: 439-448.
- Pathare, V. S., K. Y. Crous, J. Cooke, D. Creek, O. Ghannoum, and D. S. Ellsworth, 2017: Water availability affects seasonal CO₂-induced

- photosynthetic enhancement in herbaceous species in a periodically dry woodland. *Glob. Change Biol.* **23** : 5164-5178.
- Prasad, P. V. V., S. R. Pisipati, R. N. Mutava, and M. R. Tuinstra, 2008: Sensitivity of grain sorghum to high temperature stress during reproductive development. *Crop Sci.* 48: 1911-1917.
- Prasad, P. V. V. and M. Djanaguiraman, 2014: Response of floret fertility and individual grain weight of wheat to high temperature stress: sensitive stages and thresholds for temperature and duration. *Funct. Plant Biol.*, 41: 1261-1269.
- Poland, J. A., P. J. Brown, M. E. Sorrells, and J. L. Jannink, 2012: Development of high-density genetic maps for barley and wheat using a novel two-enzyme genotyping-by-sequencing approach. *PLoS ONE* 7: e32253.
- Poland, J. A. and T. W. Rife, 2012: Genotyping-by-Sequencing for Plant Breeding and Genetics. *Plant Genome* **5**: 92-102.
- Rang, Z. W., S. V. K. Jagadish, Q. M. Zhou, P. Q. Craufurd, and S. Heuer, 2011: Effect of high temperature and water stress on pollen germination and spikelet fertility in rice. *Environ. Exp. Botany* **70**: 58-65.
- Sage, R. F. and D. S. Kubien, 2007: The temperature response of C_3 and C_4 photosynthesis. Plant *Cell Environ.* 30: 1086-1106.
- Shan, Q., Y. Wang, J. Li, and C. Gao, 2014: Genome editing in rice and wheat using the CRISPR/Cas system. *Nat. Protoc.* **9**: 2395-2410.
- Shi, J., H. Gao, H. Wang, H. R. Lafitte, R. L. Archibald, and M. Yang, 2017: ARGOS8 variants generated by CRISPR-Cas9 improve maize grain yield under field drought stress conditions. *Plant Biotechnol. J.* **15**: 207-216.
- Silva, G., L. Poirot, R. Galetto, J. Smith, G. Montoya, and P. Duchateau, et al. 2011: Mega nucleases and other tools for targeted genome engineering: perspectives and challenges for gene therapy. *Curr. Gene Ther.* 11: 11-27.
- Sinsawat, V., J. Leipner, P. Stamp, and Y. Fracheboud, 2004 : Effect of heat stress on the photosynthetic apparatus in maize (*Zea mays* L.) grown at control or high temperature. *Environ. Exp. Botany* **52**: 123-129.
- Singh, B., S. N. Dheeravathu, and K. Usha, 2010: Micronutrient Deficiency: A Global Challenge and Physiological Approach to Improve Grain Productivity under Low Zinc Availability, In: Plant stress. Global science book UK, (Special issue -2) 4: 76-93.
- Singh, K., S. N. Dheeravathu, P. W. Ramteke, Reetu, N. Dikshit, and T. B. Vadithe, 2020: Effect of salt stress on morpho-physiological and green fodder yield of Bajra Napier Hybrids and TriSpecific Hybrid. *Forage Res.* 46: 241-247.

- Singh, T., S. N. Dheeravathu, N. Dikshit, N. Manjunatha, and G. Sahay, 2021: Collection and evaluation of genetic diversity in Dinanath grass (*Pennisetum pedicellatum* Trin.) for forage yield and leaf blight resistance. *J. Environ. Biol.* 42:1355-1362.
- Srivastava, R. K., R. B. Singh, V. L. Pujarula, S. Bollam, M. Pusuluri, T. S. Chellapilla, R. S. Yadav, and R. Gupta, 2020: Genome-Wide Association Studies and Genomic Selection in Pearl Millet: Advances and Prospects. *Front. Genet.* **10**: 1389.
- Takeoka, Y., K. Hiroi, H. Kitano, and T. Wada, 1991: Pistil hyperplasia in rice spikelets as affected by heat-stress. *Sexual Plant Reprod.* **4**: 39-43.
- Thirunavukkarasu, N., F. Hossain, and K. Arora, 2014: Functional mechanisms of drought tolerance in subtropical maize (*Zea mays* L.) identified using genome-wide association mapping. *BMC Genomics* 15: 1182.
- Townsend, J. A., D. A. Wright, R. J. Winfrey, F. Fu, M. L. Maeder, and J. K. Joung, 2009: High-frequency modification of plant genes using engineered zinc-finger nucleases. *Nature* **459**: 442-445.
- Varshney R. K., C. Shi, M. Thudi, C. Mariac, J. Wallace, and P. Qi et al. 2017: Pearl millet genome sequence provides a resource to improve agronomic traits in arid environments. *Nat. Biotechnol.* **35**: 969.
- Xu, Y., and J. H. Crouch, 2008: Marker-assisted selection

- in plant breeding: From publications to practice. *Crop Sci.* **48** : 391-407.
- Xue, Y., M. L. Warburton, M. Sawkins, X. Zhang, T. Setter, and Y. Xu, et al. 2013: Genome-wide association analysis for nine agronomic traits in maize under well-watered and water-stressed conditions. *Theor. Appl. Genet.* **126**: 2587-2596.
- Yadav, O. P., E. Weltzien-Rattunde, and F. R. Bidinger, 2003: Genetic variation in drought response of landraces of pearl millet (Pennisetum glaucum (L.) R. Br.). *Indian J. Genet.* **63**: 37-40.
- Yadav, R. S., C. T. Hash, F. R. Bidinger, G. P. Cavan, and C. J. Howarth, 2002: Quantitative trait loci associated with traits determining grain and stover yield in pearl millet under terminal drought-stress conditions. *Theor. Appl. Genet.* **104**: 67-83.
- Yuan, Y., J. E. Cairns, R. Babu, M. Gowda, and D. Makumbi, 2019: Genome-Wide Association Mapping and Genomic Prediction Analyses Reveal the Genetic Architecture of Grain Yield and Flowering Time Under Drought and Heat Stress Conditions in Maize. Front. Plant Sci. 9: 1919.
- Zhang, H., J. Zhang, P. Wei, B. Zhang, F. Gou, and Z. Feng, 2014: The CRISPR/Cas9 system produces specific and homozygous targeted gene editing in rice in one generation. *Plant Biotechnol. J.* 12: 797-807.